



**UNIVERSIDADE ESTADUAL DE MARINGÁ**  
**CENTRO DE CIÊNCIAS AGRÁRIAS**  
Programa de Pós-Graduação em Ciência de Alimentos

**DETERMINAÇÃO DOS COMPOSTOS QUÍMICOS,  
ATIVIDADE ANTIOXIDANTE E CITOTOXICIDADE DO  
FRUTO DA PALMEIRA *AIPHANES ACULEATA***

**LAIZA BERGAMASCO BELTRAN**

Maringá  
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Dissertação apresentada ao programa de Pós Graduação em Ciência de Alimentos da Universidade Estadual de Maringá, como parte dos requisitos para obtenção do título de mestre em Ciência de Alimentos.

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aculeata*”.**

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*Andresa Carla Feihmann*

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**Profa. Dra. Andresa Carla Feihrmann**

*Raquel Gutierrez Gomes*

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**Profa. Dra. Raquel Gutierrez Gomes**

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**Profa. Dra. Angélica Marquetotti  
Salcedo Vieira  
Orientadora**

Maringá  
2021

**Orientadora**

Dra. Angélica Marquetotti Salcedo Vieira

## **BIOGRAFIA**

Laiza Bergamasco Beltran nasceu no Paraná na cidade de Maringá. Possui graduação em Tecnologia em Alimentos pela Universidade Tecnológica Federal do Paraná. Tem experiência nas áreas de análises de alimentos e extração de compostos bioativos atuando principalmente nos seguintes temas: Avaliação da atividade antioxidante, caracterização e análises físico-químicas de alimentos.

***Dedico***

A todos que influenciaram direta e indiretamente para a conclusão deste trabalho!

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# APRESENTAÇÃO

Esta dissertação de mestrado está apresentada na forma de um artigo científico.

Laiza Bergamasco Beltran, Karine Campos Nunes, Raquel Gutierrez Gomes, Celso Vataru Nakamura, Angélica Marquetotti Salcedo Vieira, Determination of chemical composition, antioxidant activity and cytotoxicity of the *Aiphanes aculeata* palm fruit, Food Chemistry – Qualis A1.

## GENERAL ABSTRACT

**INTRODUCTION:** The Brazilian territory represents an immense heritage of natural resources, with an emphasis on exotic fruit species, many of which remain unexplored, such as potential sources for the food industry. The fruits of Arecaceae palms are considered rich in bioactive compounds, especially carotenoids, tocopherols, vitamin C and vitamin A. They also contain phenolic compounds, fibers and minerals. However, the trade of native Brazilian species is not very representative and most palms are exclusively destined for the local and regional market or are grown specifically for ornamental and landscape purposes. Therefore, the discovery and knowledge of the bioactive compounds content and antioxidant capacity of native fruits aims to give commercial and industrial value to these fruits, whose potential is still unexplored, in addition to contributing to the conservation of the Brazilian biome.

**AIMS:** Due to its little-known potential, this study aimed to analyze the composition, physicochemical properties, micronutrients, antioxidant activity and cytotoxicity of the pulp of *Aiphanes aculeata* palm.

**MATERIALS AND METHODS:** Fruits were harvested directly from several palms between June and September at the State University of Maringá, UEM/PR, Brazil. After harvesting, fruits were sorted and cleaned, packed in plastic bags and stored in a freezer at -18 °C until analysis. To obtain the pulp, the mesocarp (pulp) and epicarp (peel) were ground in a blender until completely homogenized. The average weight of the fruit was 2.5 g, and the quantity required to perform all analyzes was 500 g on a wet basis. The following analyzes were performed to determine the proximate composition: Moisture, ash, protein, lipids, fiber and carbohydrates. The physicochemical parameters were determined by analyzing the pH, soluble solids, titratable acidity, and reducing and non-reducing sugars. In the analysis of micronutrients, the mineral elements calcium (Ca), magnesium (Mg), potassium (K), iron (Fe) and phosphorus (P) were determined by atomic absorption spectrophotometric method. Vitamin C was determined by titration with potassium iodate. For the analysis of antioxidant activity and total phenols, the bioactive compounds were extracted using water, ethyl alcohol, methyl alcohol and acetone as solvents. The aqueous extract was carried out at three temperatures: 25, 40 and 90 °C. Antioxidant activity was determined by DPPH, FRAP and ABTS method and total phenolic content by Folin-Ciocalteu method. Cytotoxicity was evaluated by MTT method in non-tumor (L-929) and tumor cells (Caco-2).

**RESULTS AND DISCUSSION:** In terms of macronutrients analyzed, moisture content was 77.13 %, ash 0.51 %, lipids 0.44 %, proteins 1.03 %, crude fiber 2.52 % and carbohydrates 18.37 %, of which 11.14 % were reducing sugars and 8.06 % were non-reducing. The pH of the fruit was in the range of 5.01, which is considered as a medium acidity fruit as reported in the literature. A value of 0.38 g citric acid/100 g of the fruit was determined for titratable acidity. In the literature, citric acid values are reported between 0.08 and 1.95 %, which are good for consumption. Of the micronutrients analyzed, potassium had the highest value (993.04 mg/100 g). Potassium-rich fruits and vegetables are considered a non-pharmacological source for the prevention and control of hypertension. The other mineral elements analyzed follow in descending order: Ca (26.61 mg/100 g), Mg (18.3 mg/100 g), P (15.2 mg/100 g), Zn (0.30 mg/100 g), Fe (0.07 mg/100 g). Consumption of vitamin C is of great importance as it is not produced by the body itself and is essential for its proper functioning. Therefore, it is necessary to take it in with food, fruits and vegetables being the main sources. The recommended

daily allowance of vitamin C for an adult is 45 mg. The pulp of *Aiphanes aculeata* contained 46.97 mg/100 g of vitamin C. With respect to the extractions performed, the acetonic and aqueous extracts at 90 °C showed greater antioxidant activity by the methods studied (DPPH, FRAP and ABTS), indicating a positive correlation with the content of total phenols (Folin-Ciocalteu). The extracts showed no cytotoxic activity in the non-tumor cell line (L-929) and there was a decrease in cell viability in the tumor line (Caco-2) above a concentration of 500 µg/mL.

**CONCLUSION:** The present study proved to be very important as the pulp of *Aiphanes aculeata* palm is poorly studied and has a rich and diverse nutrient composition. The extracts showed no cytotoxic activity in non-tumor cells (L-929) at the concentrations studied, which is a positive indicator for their use in food formulations with functional properties. Thus, the results obtained show the importance of research and knowledge of native species, as they are a source of bioactive compounds and have great nutritional and economic potential.

**Keywords:** *Aiphanes aculeata*, Brazilian fruits, Bioactive compounds, Antioxidant activity, Cytotoxicity.

## RESUMO GERAL

**INTRODUÇÃO:** O território brasileiro constitui um patrimônio imensurável de recursos naturais, com ênfase em espécies de frutas exóticas, sendo muitas ainda inexploradas, as quais são fontes em potencial para a indústria de alimentos. Os frutos das palmeiras *Arecaceae* são considerados ricos em compostos bioativos, principalmente carotenoides, tocoferóis, vitamina C e vitamina A. Além disso, possuem considerável teor de compostos fenólicos, fibras e minerais. No entanto, o comércio de espécies nativas brasileiras é pouco representativo e a maioria das palmeiras são destinadas exclusivamente ao mercado local e regional ou cultivada especificamente para fins ornamentais e paisagísticos. Portanto, a descoberta e o conhecimento do conteúdo de compostos bioativos e da capacidade antioxidante de frutas nativas visa agregar valor comercial e industrial a estas frutas que possuem seu potencial ainda inexplorado, além de contribuir com a conservação do bioma brasileiro.

**OBJETIVO:** Devido seu potencial ainda pouco conhecido, este estudo teve como objetivo realizar as análises de composição centesimal, físico-químicas, micronutrientes, atividade antioxidante e citotoxicidade da polpa do fruto da palmeira *Aiphanes aculeata*.

**MATERIAIS E MÉTODOS:** Os frutos foram coletados entre os meses de junho e setembro diretamente de diversas palmeiras localizadas na Universidade Estadual de Maringá, UEM/PR, Brasil. Após a colheita, procedeu-se com a seleção e higienização dos frutos e estes foram embalados em sacos plásticos e acondicionados em freezer à -18 °C até o momento da realização das análises. Para a obtenção da polpa, o mesocarpo (polpa) e epicarpo (casca) foram triturados em liquidificador até a total homogeneização. O peso médio dos frutos é 2,5 g e a quantidade necessária para realização de todas as análises foi de 500 g em base úmida. Realizou-se as seguintes análises para determinar a composição centesimal: Umidade, cinzas, proteína, lipídios, fibra e carboidratos. Os parâmetros físico-químicos foram determinados através das análises: pH, sólidos solúveis, acidez titulável, açúcar redutor e não redutor. Em relação a análise de micronutrientes os elementos minerais cálcio (Ca), magnésio (Mg), potássio (K), ferro (Fe) e fósforo (P) foram determinados utilizando a técnica espectrofotométrica de absorção atômica. A vitamina C foi determinada por titulação com iodato de potássio. Para as análises de atividade antioxidante e fenólicos totais, foi realizada a extração de compostos bioativos utilizando os solventes: água, álcool etílico, álcool metílico e acetona. O extrato aquoso foi realizado em três temperaturas 25, 45 e 90 °C. A atividade antioxidante foi realizada através dos métodos DPPH, FRAP e ABTS e o conteúdo fenólico total foi determinado utilizando o método de Folin-Ciocalteu. A citotoxicidade foi avaliada pelo método MTT em células não-tumorais (L-929) e tumorais (Caco-2).

**RESULTADOS E DISCUSSÃO:** Em relação aos macronutrientes analisados obteve-se um teor de umidade de 77,13 %, cinzas 0,51 %, lipídeos 0,44 %, proteínas 1,03 %, fibra bruta 2,52 % e carboidratos 18,37 %, sendo 11,14 % de açúcar redutor e 8,06 % de não redutor. Em relação ao pH do fruto, este se apresentou na faixa de 5,01, sendo considerado uma fruta de acidez média conforme proposto pela literatura. Quanto a acidez titulável o valor obtido foi de 0,38 g ácido cítrico/100 g de fruta. Valores de ácido cítrico em frutas variando de 0,08 a 1,95 % é reportado na literatura que são bem aceitos para consumo. Dentre os micronutrientes analisados o potássio apresentou maior quantidade (993,04 mg/100 g). Frutas e vegetais ricos em potássio são considerados fontes

não-farmacológicas para a prevenção e controle da hipertensão. Segue em ordem decrescente os demais elementos minerais analisados: Ca (26,61 mg/100 g), Mg (18,3 mg/100 g), P (15,2 mg/100 g), Zn (0,30 mg/100 g), Fe (0,07 mg/100 g). O consumo de vitamina C é de grande importância, pois ela não é naturalmente produzida pelo organismo sendo imprescindível para seu bom funcionamento. Deste modo, é necessário ingeri-la, tendo as frutas e hortaliças como suas principais fontes. A recomendação diária de vitamina C para um adulto é de 45 mg. A polpa do fruto da *Aiphanes aculeata* apresentou 46,97 mg/100 g de vitamina C. Em relação aos extratos realizados, o acetônico e aquoso a 90 °C apresentaram maior atividade antioxidante nas metodologias estudadas (DPPH, FRAP e ABTS), indicando correlação positiva com o teor de fenólicos totais (Folin-Ciocalteu). Os extratos não apresentaram atividade citotóxica na linhagem de células não tumorais (L-929) e houve diminuição da viabilidade celular na linhagem tumoral (Caco-2) a partir da concentração de 500 µg/mL.

**CONCLUSÃO:** O presente estudo se mostrou de grande importância, pois a polpa do fruto da palmeira *Aiphanes aculeata* é ainda pouco explorada e apresentou uma composição nutricional rica e variada. Os extratos não apresentaram atividade citotóxica nas concentrações estudadas em células não tumorais (L-929), o que é um indicador positivo para ser usado em formulações alimentícias com propriedades funcionais. Os resultados obtidos demonstram a necessidade de pesquisas e conhecimento sobre as espécies nativas, visto que são fonte de compostos bioativos e apresentam grande potencial nutricional e econômico.

**Palavras-chave:** *Aiphanes aculeata*, frutas brasileiras, compostos bioativos, atividade antioxidante, citotoxicidade.



43 phenolic compounds (De Souza, Araújo, Farias, Zanotto, Neri-Numa & Pastore, 2020).  
44 However, the trade of native Brazilian species is not very representative, and most palms are  
45 exclusively destined for the local and regional market or are grown specifically for ornamental  
46 and landscape purposes (Campos, De Lima Araújo, Gaoue, & Albuquerque, 2019).

47         Recently, studies have shown that several fruits of the Arecaceae family have important  
48 biological activities, such as anti-inflammatory properties (*Cocos nucifera* L.), preventive  
49 effect on hypoglycemia (*Syagrus romaziffiana*) (Ribeiro, Bieski, Balogun, & Martins, 2017)  
50 and photochemoprotective effect (oil from the pulp of *Mauritia flexuosa*) (Zanatta, Mitjans,  
51 Urgatondo, Rocha-Filho, & Vinardell, 2010). However, no data on the nutrient composition  
52 and antioxidant potential of the palm fruits of *Aiphanes aculeata* were found in the literature.

53         Fruits are generally considered to be a rich source of nutrients. Several studies have  
54 found a positive correlation between phenolic compound content and antioxidant potential  
55 (Santos-Buelga, González-Paramás, Oludemi, Ayuda-Durán, & González-Manzano, 2019). A  
56 high consumption of foods with phenolic compounds in their composition has been associated  
57 with a lower incidence of diseases such as inflammation, cancer, cardiovascular dysfunction,  
58 and a decrease in the immune system. In addition, they play an important role in the prevention  
59 of neurodegenerative diseases and antiviral activity (Liu, Wang, & Wang, 2018).

60         The antioxidant activity of phenolic compounds is attributed to their ability to scavenge  
61 free radicals by donating hydrogen atoms, electrons, or chelating metal cations, thus inhibiting  
62 the initiation or even the continuation chain of free radical-induced oxidative reactions in the  
63 human organism. The action of these compounds is thus associated with the attenuation of  
64 oxidative processes, which may contribute to the protection of body cells from damage by  
65 providing a protective effect against reactive oxygen species (Santos-Buelga, González-  
66 Paramás, Oludemi, Ayuda-Durán, & González-Manzano, 2019).

67 The discovery and knowledge of the bioactive compounds content and antioxidant  
68 capacity of indigenous fruits aims to give commercial and industrial value to these fruits, which  
69 have an unexplored potential. In addition, it could contribute to the conservation of the  
70 Brazilian biome by providing nutritional alternatives and reducing the harmful effects of free  
71 radicals due to their antioxidant properties (Teixeira et al. 2019). With this in mind, this work  
72 aimed to perform analyzes of proximate composition, physicochemical properties,  
73 micronutrients, antioxidant activity and cytotoxicity of *Aiphanes aculeata* palm fruit, since its  
74 potential has not yet been explored.

75

## 76 2. Materials and methods

### 77 2.1 Chemical reagents

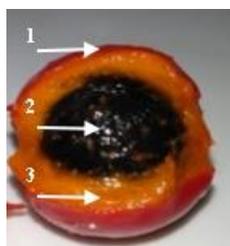
78 The respective SYNTH brand reagents were used: hexane, acetone, ethyl alcohol,  
79 methyl alcohol, acetone, sulfuric acid, boric acid, hydrochloric acid, gallic acid,  
80 dimethylsulfoxide (DMSO), potassium sulfate, phenolphthalein, potassium iodide , potassium  
81 iodate, starch, sodium carbonate. The respective DYNAMIC brand reagents were used: copper  
82 sulfate, methyl red, sodium hydroxide, fehling solution A and B, methylene blue. The  
83 respective SIGMA-ALDRICH brand reagents were used: ABTS (2,2'-azino- bis-(3-  
84 ethylbenzenethiazoline-6-sulfonic acid), DPPH (2,2-diphenyl-1-picrylhydrazyl), trolox (6-  
85 hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid), TPTZ (2,4,6-Tri-(2-Pyridyl)-1,3,5-  
86 Triazine 1:3 with p-Toluenesulfonic acid), Folin-Ciocalteu reagent, quercetin, 3-(4,5-  
87 dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT). The respective FBS - Life  
88 Technologies/Gibco Laboratories brand reagents were used: Dulbecco's Modified Eagle's  
89 medium (DMEM); fetal bovine serum.



90 **Fig. 1.** *Aiphanes aculeata* palm trees located at the State University of Maringá.  
 91  
 92

### 93 2.2 *Aiphanes aculeata* palm tree fruit

94 The fruits were collected between June and September, 2019 (Figure 1), directly from  
 95 the palm trees located at the State University of Maringá (UEM), Paraná, Brazil, (-23.4051563,  
 96 -51.93876817195696). After harvesting, fruits selection and cleaning were carried out, being  
 97 further packaged in plastic bags and stored in a freezer at -18 °C until the analysis performance.



98 **Fig. 2.** Overview of *Aiphanes aculeata* fruit; 1. epicarp; 2. endocarp; 3. mesocarp.  
 99  
 100

### 101 2.3 Fruit pulp obtaining

102 To obtain the fruit pulp used in the analysis, we proceeded with the fruits thawing and  
 103 pulping, in which the epicarp and mesocarp (Figure 2) were placed in a blender until (Britânia  
 104 700w) total homogenization. The average weight of the fruits was 2.5 g and the amount needed  
 105 to carry out all analyses was 500 g of the pulp *in natura*.

### 107 2.4 Proximate composition analysis

108           The following analyses were performed in triplicate. Moisture and ash contents were  
109 determined by drying in an oven with circulating air at 105 °C (934.06 - AOAC, 2016) and  
110 incineration in a muffle at 550 °C (940.26 - AOAC, 2016) respectively; crude protein was  
111 determined by the Kjeldahl method (960.52 - AOAC, 2016), using the conversion factor 5.75  
112 for vegetable proteins according to the RDC ANVISA Resolution No 360, of December 23,  
113 2003; the lipid content was performed by Soxhlet extraction using hexane as an extracting  
114 solvent (920.39 - AOAC, 2016). Crude fiber analysis was determined by acid hydrolysis  
115 following the method (958.06 - AOAC, 2005). Carbohydrate was calculated by difference  
116 using the formula:  $100 - (\text{moisture} + \text{lipid} + \text{protein} + \text{ash} + \text{crude fiber})$ .

117

#### 118 *2.5 Physicochemical analysis*

119           The pulp was evaluated for pH using a calibrated digital potentiometer (AOAC, 1992);  
120 soluble solids were determined by direct reading in a bench-top refractometer (Abbe, 2WAJ)  
121 (AOAC, 1992); determination of the titratable acidity, this method is based on the  
122 potentiometric titration of the sample with a 0.1 M sodium hydroxide solution (942.15b –  
123 AOAC, 1995); the content of reducing and non-reducing sugars was determined using the  
124 Fehling titration technique, and the results were expressed in percentages of glucose and  
125 sucrose, respectively (958.06 – AOAC, 1995). All analyses were performed in triplicate.

126

#### 127 *2.6 Micronutrient Analysis*

128           The mineral micronutrients analyses were carried out after the solubilization of the  
129 fixed mineral residue. It was conducted according to the method No 975.03 described by the  
130 Association of Official Analytical Chemists (AOAC, 2005), using the spectrophotometric  
131 technique of atomic absorption (Varian, Spectra AA 50B), which analyzed the calcium (Ca),  
132 magnesium (Mg), potassium (K), iron (Fe) and phosphorus (P) mineral elements.

## 133 2.7 Vitamin C analysis

134 Vitamin C content was determined by the procedure described by Association of  
135 Official Analytical Chemists, method 43.064 (AOAC, 1992). The method used was based on  
136 the vitamin C oxidation by titration with potassium iodate. The analyses were performed in  
137 triplicate and the results were expressed in mg of pulp vitamin C/100g.

138

## 139 2.8 Extraction of bioactive compounds

140 For the analysis of antioxidant activity and total phenolics, bioactive compounds were  
141 extracted using the following solvents: water, ethyl alcohol, methyl alcohol and acetone. All  
142 extractions were performed following the methods proposed by Roesler et al. (2007), with  
143 some modifications.

144

### 145 2.8.1 Aqueous extract

146 As regards the aqueous extract, the pulp was homogenized for 20 minutes with distilled  
147 water at a ratio of 1: 3 (m/m) pulp: water in a magnetic stirrer (MS-H-Pro) at 420 rpm. The  
148 material was filtered with a Whatman No1 paper filter and the residue was re-extracted with  
149 water under the same conditions. This procedure was performed at 25, 45 and 90 °C. The  
150 extracts obtained were lyophilized (Alpha 1-4 LO plus) and stored in amber vials at -18 °C.

151

### 152 2.8.2 Ethanol, methanol and acetonic extract

153 In an Erlenmeyer flask, the pulp was added at a ratio of 1:3 (m/m) pulp: extracting  
154 solution for both solvents with a concentration of 3:7 (v/v) water: solvent. The sample was  
155 homogenized for 20 minutes at room temperature ( $25 \pm 2$  °C) in a magnetic stirrer (MS-H-Pro)  
156 at 420 rpm and filtered with a Whatman No. 1 paper filter. The residue was washed with 30  
157 mL of extractor solution. The filtrate was rota-evaporated (Tecnal TE-211) at 50 °C to recover

158 the solvent. The extracts obtained were lyophilized (Alpha 1-4 LO plus) and stored in amber  
159 vials at -18 °C.

160

## 161 *2.9 Antioxidant capacity evaluation*

### 162 *2.9.1 DPPH· (free radical scavenging) assay*

163 The ability of extracts to scavenge the DPPH radical was evaluated according to the  
164 methods proposed by Rufino et al. (2007), with some modifications. To prepare the DPPH  
165 reagent, 1 mg of DPPH was diluted in 1 mL of methanol, obtaining a solution with a  
166 concentration of 130 µM. The extracts were diluted in different concentrations (1000 - 31  
167 µg/mL). Thereafter, 100 µl of each dilution in the 96-well transparent microplate and 100 µl of  
168 the 130 µM DPPH solution were added. The microplate was incubated at room temperature  
169 protected from light for 30 minutes. The reading was performed at 517 nm in a microplate  
170 reader (BioTek, Power Wave XS). Antioxidant capacity was expressed as the sample  
171 concentration needed to reduce the initial number of free radicals by 50 % (EC<sub>50</sub>). The results  
172 were expressed in µg/mL.

173

### 174 *2.9.2 Ferric reducing antioxidant power (FRAP) assay*

175 The following solvents were used to prepare the FRAP reagent: 0.3 mM of acetate  
176 buffer (pH 3.6), TPTZ 10 mM reagent in HCl 40 mM and FeCl<sub>3</sub> 20 mM solution. These  
177 solutions were mixed in a 10:1:1 ratio, resulting in the final solution of the FRAP reagent. In a  
178 96-well transparent microplate, 30 µL of the previously prepared extracts (1000 – 31 µg/mL)  
179 and 150 µL of the prepared FRAP reagent were added. This solution was incubated for 30  
180 minutes at 37 °C in an oven with air circulation. After stabilization at room temperature, the  
181 reading was performed at 595 nm in a microplate reader (BioTek, Power Wave XS). Trolox  
182 was used as the standard antioxidant, and the results were expressed as micromoles of Trolox

183 equivalents per gram of sample ( $\mu\text{mol}$  of TE / g sample). The test was carried out using the  
184 methods proposed by Rufino et al. (2010) with modifications.

185

### 186 2.9.3 *ABTS $\cdot^+$ assay*

187 The capacity of the extracts to scavenge the cationic radical  $\text{ABTS}\cdot^+$  was evaluated in  
188 accordance with the procedures proposed by Rufino et al. (2010), with some modifications.

189 The  $\text{ABTS}\cdot^+$  radical was obtained by mixing 5 mL of the ABTS solution (7.0 mmol/L) and 88  
190  $\mu\text{L}$  of the potassium persulfate solution (2.45 mmol/L) for 12 h, in the absence of light.

191 Subsequently, ethyl alcohol was added to this solution up to an absorbance of  $0.700 \pm 0.05$  at

192 754 nm in a microplate reader. In a 96-well transparent microplate, 7  $\mu\text{L}$  of the previously

193 prepared dilutions (1000 – 31  $\mu\text{g}/\text{mL}$ ) and 200  $\mu\text{L}$  of the ABTS solution were added, and the

194 mixture was homogenized. After 6 minutes of reaction, the reading was performed at 734 nm

195 in a microplate reader (BioTek, PowerWave XS). Trolox was used as the standard antioxidant.

196 The results were expressed as micromoles of Trolox equivalents per gram of sample ( $\mu\text{mol}$  of  
197 TE / g of sample).

198

### 199 2.9.4 *Total Phenolics determination*

200 The total phenolic content was determined according to the methods proposed by Pires

201 et al. (2017) with modifications. Accordingly, 20  $\mu\text{L}$  of the previously prepared dilutions (5 -

202 0.31 mg/mL) were added in the following solutions: 180  $\mu\text{L}$  of deionized water, 20  $\mu\text{L}$  of

203 methanol, 20  $\mu\text{L}$  of Folin-Ciocalteu reagent (1N) and 60  $\mu\text{L}$  of carbonate solution sodium (10

204 %) in a 96-well clear microplate. The microplate was incubated for 20 minutes at room

205 temperature and protected from direct light. The reading was performed at 760 nm in a

206 microplate reader (BioTek, PowerWave XS). The results were expressed in mg of equivalents

207 in gallic acid (GAE) per 100 g of fresh fruit (mg GAE/100 g of fresh fruit).

## 2.10 Cell viability assay

L-929 fibroblasts (ATCC®CCL-1.3™) and human colon adenocarcinoma cell line Caco-2 (ATCC, #HTB-37) were cultured in DMEM with 10 % fetal bovine serum (FBS) and 1 % penicillin-streptomycin in a 5 % atmospheric CO<sub>2</sub> at 37 °C. The cultured cell suspension was plated in a 96-well plate (2.5 × 10<sup>5</sup>cells/well) and incubated for 24 hours. Afterwards, the medium was removed and the cells were treated with different samples concentrations: 31.25-250 µg/mL for L-929 and 31.25-500 µg/mL for Caco-2. After 24 hours, the cells were washed with PBS and received 50 µL of MTT (2 mg/mL), being incubated for 4 hours. Finally, 200 µL of DMSO were added and the absorbance was determined at 570 nm (BioTek, PowerWave XS microplate spectrophotometer) (Berridge, Herst & Tan, 2005)

## 2.11 Statistical analysis

Statistical analysis was performed using GraphPad Prism software v. 5.00 (San Diego, CA, USA). Data were analyzed using the ANOVA (one-way or two way), followed by Tukey post hoc or Bonferroni correction, considering  $p \leq 0.05$  as statistically significant. Data were expressed as mean ± standard deviation of at least three independent experiments.

## 3. Results and discussion

Table 1 shows the results of the analyses of proximate composition, physicochemical characterization, and Vitamin C of the pulp of the *Aiphanes aculeata* palm fruit.

**Table 1**  
Results of the proximate composition, physicochemical characterization and Vitamin C analyses of the *Aiphanes aculeata* fruit pulp.<sup>a</sup>

Analyses	<i>Aiphanes aculeata</i> pulp <i>in natura</i>
Moisture (%)	77.13 ± 0.22
Ash (%)	0.51 ± 0.01
Lipids (%)	0.44 ± 0.04
Protein (%)	1.03 ± 0.20

Fiber (%)	2.52 ± 0.01
Carbohydrates (%)	18.37 ± 0.09
pH	5.01 ± 0.04
Soluble solids (° Brix)	18.42 ± 0.03
Reducing sugar (%)	11.14 ± 0.68
Non-reducing sugar (%)	8.06 ± 0.30
Titrateable Acidity (g acid citric /100 g)	0.38 ± 0.09
Vitamin C (mg/100 g of fresh pulp)	46.97 ± 5.08

<sup>a</sup> Mean value ± standard deviation based of fruit pulp weight; n=3.

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Moisture content is a measure of the total amount of water in the food. It is usually expressed as a percentage from the total weight. The moisture content in the *Aiphanes aculeata* fruit (77.13 %) is in the expected range for fresh fruit since the water content can vary from 65 to 95 % (Cecchi, 2003). The moisture content found was higher than that presented by Lescano et al. (2018) for *Syagrus romanzoffiana* (69.62 %) and *Mauritia flexuosa* (73.45 %), which are fruits from the same botanical family.

The minerals or ash are obtained through the incineration process using high temperatures, burning the sample's organic matter. Thus, through the resulting content, it is possible to obtain the number of specific components such as minerals Ca, Mg, Fe, Zn, P, K, among other micronutrients. These elements are necessary for the proper functioning of vital functions, as well as for the good health of the human body (Zambiasi, 2010). The ash content found in the *Aiphanes aculeata* pulp (0.51 %) is similar to the content of other fruits from the Brazilian cerrado such as *Psidium cattleianum* (0.43 %), *Mauritia flexuosa* (1.01 %), *Stenocalyx dysentericus* (0.30 %), *Hancornia speciosa* (0.53 %) e *Annona crassiflora* (0.68 %), reported by Schiassi, Souza, Lago, Campos and Queiroz (2018).

The *Aiphanes aculeata* fruits can be considered a low source of lipids (0.44 %). When analyzing fruits from the Brazilian cerrado, Schiassi, Souza, Lago, Campos and Queiroz (2018) found a low lipid value for *Eugenia dysenterica* DC. (0.49 %) and *Spondias mombin* L. (0.48 %) fruits, with similar values from those found in the present work.

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253 The protein content found in the *Aiphanes aculeata* fruit (1.03 %) presented a value  
254 close to that described by Hamacek, Della Lucia, Silva, Moreira and Pinheiro-Sant'ana (2018)  
255 for the *Mauritia flexuosa* L pulp (1.00 %) and a higher value than that found by Becker, Chagas,  
256 Marty, Mendes and Nunes (2018) for *Maximiliana maripa* pulp (0.49 %) and *Euterpe oleracea*  
257 Mart. (0.72 %), both fruits from the Arecaceae family.

258 Regarding the total fiber content, the value found for the *Aiphanes aculeata* pulp was  
259 lower than those reported for fruits from the same species, as presented by Hameck, Della  
260 Lucia, Silva, Moreira and Pinheiro-Sant'ana (2018) for the *Mauritia flexuosa* L pulp (9.23 %),  
261 and for the *Attalea phalerata* fruit pulp (6.33 %), as reported by Lescano et al. (2018).

262 Amongst the macronutrients, fruits have a higher carbohydrate content in relation to  
263 proteins and lipids (Lescano et al. 2018). The pulp of *Aiphanes aculeata* had a higher  
264 carbohydrate content (18,37 %) in comparison to *Mauritia flexuosa* pulp from different biomes.  
265 A content of 9.70 % and 11.31 % were reported from the Cerrado and Amazonian fruits,  
266 respectively (Cândido & Silva, 2017). The sugar content in fruits is influenced by the amount  
267 of light the plants receive. As photosynthesis occurs for a longer time, it is possible to transport  
268 sugar reserves to the fruits, therefore, the greater the degree of maturation of the fruit, the  
269 greater its sweetness (Taiz & Zeiger, 2006).

270 According to the classification proposed by the Food and Drug Administration (FDA,  
271 2016), the *Aiphanes aculeata* fruits are classified as medium acidity fruits (pH 5.01), in which  
272 the pH ranges from 4.6 to 5.3. Acidic fruits have a pH range between 3.7 and 4.6 and very  
273 acidic fruits have a pH < 3.7. For the *Mauritia vinifera* fruit pulp, which is from the same  
274 family as *Aiphanes aculeata*, Hamacek, Della Lucia, Silva, Moreira and Pinheiro-Sant'ana  
275 (2018) reported a pH of 3.59.

276 The soluble solids content relates to the total sugar content, an important aspect when  
277 correlated with the sweetness of the fruit, since sweet fruits are usually more accepted by

278 consumers. The *Aiphanes aculeata* pulp had a total solids content of 18.42 °Brix. In view of  
279 this, De Souza et al. (2020) reported total solids contents for the *Mauritia flexuosa* (13.40  
280 °Brix), *Euterpe oleracea* Mart. (6.46 °Brix) and *Acrocomia aculeata* (29.70 °Brix), all fruits  
281 from the Arecaceae family. Thus, the *Aiphanes aculeata* fruit has a high content of soluble  
282 solids, being superior to several fruits from the same botanical family, which are already known  
283 and accepted by consumers.

284         Regarding titratable acidity, the value obtained for the *Aiphanes aculeata* fruit (0.38 g  
285 citric acid/100 g) was lower than those reported by Hamacek, Della Lucia, Silva, Moreira and  
286 Pinheiro-Sant'ana (2018) for the *Mauritia vinífera* fruit pulp (1.42 g citric acid/100 g), which  
287 also belongs to the Arecaceae family.

288         One of the most important quality factors of fruits is the flavor, assessed by the content  
289 of soluble sugars and organic acids in it. During ripening, the organic acid content decreases  
290 and the sugar content increases. The reducing sugar refers to the glucose content and the non-  
291 reducing sugar represents the sucrose content in the fruit (Batista-Silva et al. 2018). Regarding  
292 the content of reducing and non-reducing sugars, values of 11.14 % and 8.06 % were found for  
293 the *Aiphanes aculeata* fruit pulp, respectively. The contents of reducing and non-reducing  
294 sugars differ among the fruits as they depend on the interaction between several variables, such  
295 as age, climatic conditions, maturation and cultivation.

296         Ascorbic acid is a water-soluble vitamin, essential for collagen synthesis and tissue  
297 repair. It plays a significant role in the metabolism of carbohydrates and in the synthesis of  
298 lipids and proteins. Additionally, it supports the immune system by means of its antioxidant  
299 property, helping to neutralize free radicals in cells. The consumption of vitamin C is of great  
300 importance, as it is not naturally produced by the human body, being essential for its proper  
301 functioning. Thus, it is necessary to acquire it by ingesting fruits and vegetables since they are  
302 its main sources (Garcia, Lima & Bomfim, 2017). According to the Dietary Reference Intakes

303 (DRIs) (2000), the recommended daily intake of vitamin C is 45 mg for adults. Therefore, a  
 304 100 mg portion of *Aiphanes aculeata* fruit provides the recommended amount of vitamin C.  
 305 The value reported for the *Aiphanes aculeata* fruit (46.97 mg/100g) was higher than those from  
 306 the *Mauritia flexuosa* L. palm tree (17.4 mg/100g) (Morais et al. 2019).

307 Fruits and vegetables are important sources of nutrients. Minerals play a vital role in  
 308 the development and proper functioning of the human body. Table 2 shows the results obtained  
 309 in the mineral composition analyzes (Ca, Mg, Zn, Fe, P and K) of the *Aiphanes aculeata* fruit  
 310 pulp. From these results, we were able to calculate the mineral contribution considering the  
 311 Dietary Reference Intake (DRI) for a healthy adult, as a percentage (%) per 100g of pulp  
 312 (Institute of Medicine, 1999-2011).

313 **Table 2**  
 314 Mineral contents and contribution according to the Dietary Reference Intake (DRI)  
 315 per 100 g of fruit pulp.<sup>a</sup>

Minerals	<i>Aiphanes aculeata</i> pulp in natura
Ca (mg/100 g)	26.61 ± 0.22
% DRI	2.67
Mg (mg/100 g)	18.3 ± 0.14
% DRI	7.07
Zn (mg/100 g)	0.30 ± 0.10
% DRI	4.28
Fe (mg/100 g)	0.07 ± 0.16
% DRI	0.50
P (mg/100 g)	15.2 ± 0.15
% DRI	2.17
K (mg/100 g)	993.04 ± 0.11
% DRI	21.13

316 <sup>a</sup> Mean value ± standard deviation; n = 3. DRI (Dietary Reference Intake), Ca  
 317 (Calcium), Mg (Magnesium), Zn (Zinc), Fe (Iron), P (Phosphorus), K (Potassium).  
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319 It follows in decreasing order the content of minerals obtained in 100 grams of the pulp  
 320 of the fruit of *Aiphanes aculeata* Potassium > Calcium > Magnesium > Phosphorus > Zinc >  
 321 Iron. The consumption of foods that have several minerals in their composition is of great  
 322 importance for health, as they have important factors in the prevention of various diseases, and  
 323 fruits are potential sources of them. Among the mineral contents analyzed, potassium (933.04

324 mg/100 g) was the most abundant element present in the *Aiphanes aculeata* fruit. The value  
 325 found was higher than those reported by De Souza et al. (2020) for *Mauritia flexuosa* (183.55  
 326 mg/100 g) and *Acrocomia aculeata* (2.36 mg/100 g), fruits from the Arecaceae family.  
 327 Potassium plays an important role in intracellular and extracellular water distribution. It is an  
 328 important factor in lowering blood pressure and in cardiovascular disease. The most recent  
 329 recommendations for the detection, prevention and control of hypertension in adults  
 330 recommend increased potassium intake through fresh foods as one of the most effective non-  
 331 pharmacological measures for the prevention and treatment of hypertension (Turck et al. 2016).

332 The analyses of total phenolic compounds and antioxidant activity were performed  
 333 using different solvents, once the extraction of bioactive compounds maximum extraction is  
 334 important to evaluate the antioxidant capacity, which have different polarity. Thus, solubility  
 335 in a particular solvent is a distinctive feature of the phytochemistry present in the fruit, which  
 336 explains the lack of a universal extraction procedure (Santos, Feitosa, Rodrigues, & Santana,  
 337 2019). Table 3 shows the results of the aqueous extract subjected to temperatures of 25, 45 and  
 338 90 °C, as well as the ethanolic, methanolic and acetonetic extracts of the *Aiphanes aculeata* fruit  
 339 by the DPPH, FRAP, ABTS and Total Phenolics tests.

340 **Table 3**  
 341 Antioxidant capacity by DPPH, FRAP, ABTS and Total Phenolic methods.<sup>a</sup>

Extracts	DPPH EC <sub>50</sub> [µg/mL] <sup>b</sup>	FRAP (µmol TE/g sample)	ABTS (µmol TE/g sample)	Total Phenolics (mg GAE/100 g of fresh fruit)
Aqueous 25 °C	352.74 ± 28,73 <sup>a</sup>	10.13 ± 1.07 <sup>ad</sup>	208.79 ± 5,59 <sup>a</sup>	393.89 ± 1.34 <sup>a</sup>
Aqueous 45 °C	274.10 ± 24,54 <sup>b</sup>	13.04 ± 2.12 <sup>a</sup>	561.97 ± 1,17 <sup>b</sup>	347.45 ± 3.87 <sup>a</sup>
Aqueous 90 °C	64.20 ± 4.18 <sup>c</sup>	41.16 ± 2.84 <sup>b</sup>	712.89 ± 1,37 <sup>c</sup>	1419.44 ± 2.16 <sup>b</sup>
Acetone 70 %	43.70 ± 1.97 <sup>c</sup>	84.27 ± 3.69 <sup>c</sup>	829.09 ± 8,97 <sup>d</sup>	1074.58 ± 5.93 <sup>c</sup>
Ethanol 70 %	146.05 ± 10.19 <sup>b</sup>	15.27 ± 1.28 <sup>d</sup>	703.19 ± 1,18 <sup>b</sup>	309.63 ± 1.70 <sup>a</sup>
Methanol 70 %	157.71 ± 16.48 <sup>b</sup>	30.62 ± 5.06 <sup>b</sup>	616.54 ± 9,52 <sup>bc</sup>	344.72 ± 2.10 <sup>a</sup>

342 <sup>a</sup> Mean value ± standard deviation; n=3. Different letters in the same column indicate a significant  
 343 difference by Tukey's test (p < 0.05). GAE (gallic acid equivalents), TE (trolox equivalents).

344 <sup>b</sup> Concentration of antioxidant required to reduce the original number of free radicals by 50%.

345 In order to obtain maximum yield of phenols, several researchers have studied  
346 extraction with different solvents. Methanol, acetone, ethanol, and water are the most common  
347 solvents used for extraction due to their polar properties, which provide affinity for phenolic  
348 compounds in general (Santos, Feitosa, Rodrigues, & Santana, 2019). These compounds are  
349 associated with the adaptation and resistance mechanisms of plants to the environment and can  
350 influence the taste, technological properties, and nutritional and functional potential of fruit  
351 species (Rocha et al. 2011).

352 The aqueous extract, in which a temperature of 90 °C was used, had the highest content  
353 of total phenolics (1419.44 mg GAE/100 g of fresh fruit). Therefore, it is evident that the  
354 compounds in the *Aiphanes aculeata* fruit have greater polarity, being more water-soluble (i.e.,  
355 have greater affinity with water). The efficiency of water in the extraction process is important  
356 because it has a more environmentally friendly extraction and low toxicity compared to organic  
357 solvents. Moreover, the use of water is a good alternative for the extraction of phenolic  
358 compounds due to its safety, accessibility and low cost. Garcia-Mendoza et al (2017) observed  
359 that by increasing the temperature from 40 °C to 80 °C, the extraction of phenolic compounds  
360 from *Euterpe edulis* fruit residues increased by almost 62 %. These results indicate that high  
361 temperatures have a positive effect on the solubility of phenolic compounds, which increases  
362 the transfer of these compounds to the solvent and improves the efficiency of extraction.

363 Vasco, Ruales & Kamal-Eldin (2008) classified total phenolic content in fruits into  
364 three categories: low (100 mg GAE /100g), medium (100-500 mg GAE /100g) and high (500  
365 mg GAE /100g) for samples from fresh material. According to this classification, the palm fruit  
366 of *Aiphanes aculeata* is classified with high total phenolic content considering the aqueous and  
367 acetonetic extract at 90 °C (1074.58 mg GAE /100 g fresh fruit). The value for total phenolic  
368 content of *Aiphanes aculeata* fruit determined in the present study was higher than the values  
369 reported by Seraglio et al. (2015) for two species of *Bactris gasipaes* palm fruits, where the

370 values reported for red peach palm and yellow peach palm were 93.35 and 87.36 mg GAE /100  
371 g of fresh fruit, respectively.

372 Currently, there is no official method for determining antioxidant activity in foods  
373 because there are different mechanisms and a variety of bioactive compounds. Therefore, more  
374 than one method is used to evaluate the antioxidant activity of a compound (Lima-Neto et al.  
375 2017).

376 As for the extracts analyzed, the antioxidant activity for DPPH free radical capture was,  
377 in decreasing order, acetone > 90 °C aqueous > ethanol > methanol > 45 °C aqueous > 25 °C  
378 aqueous. According to Villaño, Fernández-Pachón, Moyá, Troncoso, & García-Parrilla (2007),  
379 the EC<sub>50</sub> value is inversely proportional to the antioxidant capacity of a compound, since the  
380 EC<sub>50</sub> value indicates the number of antioxidants required to reduce the concentration of the  
381 radical by 50 %. This means that the lower the EC<sub>50</sub> value, the greater the antioxidant activity.

382 Amongst the extracts analyzed, there was no significant difference ( $p < 0.05$ ) between  
383 acetic (43.70 µg/mL) and aqueous at 90 °C (64.20 µg/mL). Roesler et al. (2007), when  
384 evaluating the antioxidant capacity of fruits from the Brazilian cerrado, found EC<sub>50</sub> values for  
385 the aqueous and ethanol extracts of the following fruits: *Eugenia dysenterica* DC (879.93 and  
386 387.47 µg/mL), *Annona montana* (1321, 93 and 148.82 µg/ml) and *Solanum lycocarpum*  
387 (1328.98 and 182.16 µg/ml). Superior results are identified when comparing acetic and  
388 aqueous extracts at 90 °C of the *Aiphanes aculeata* fruit, since it was only necessary 43.70 and  
389 64.20 µg / mL, respectively, to capture 50 % of the radicals.

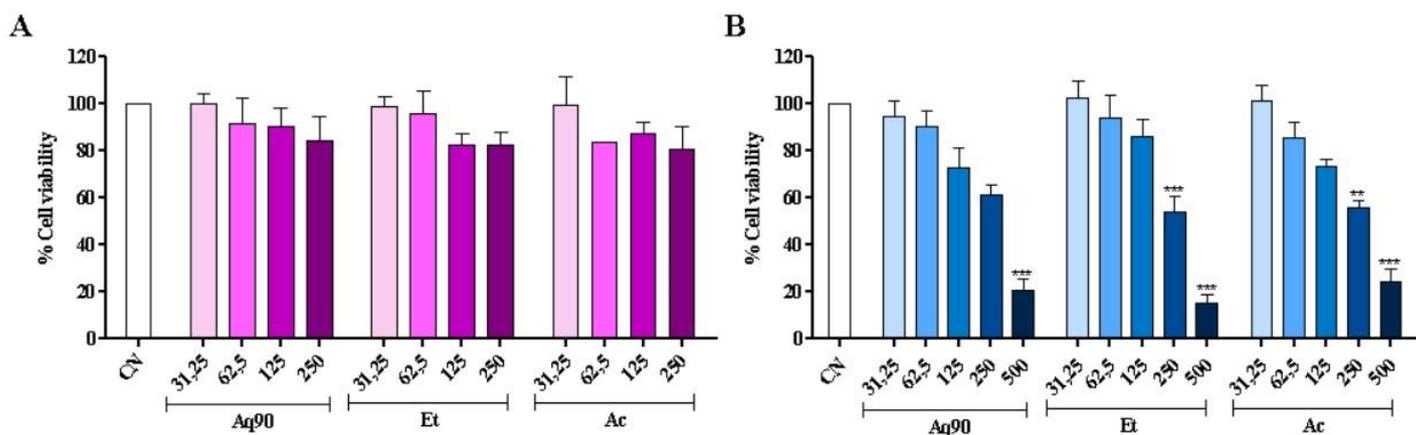
390 In terms of Fe<sup>+3</sup> reducing power, the acetone extract had the highest activity (84.24  
391 µmol TE/g sample), followed by the extracts (in decreasing order): aqueous 90 °C > methanol  
392 > ethanol > aqueous 45 °C > 25 °C aqueous. Pásko et al. (2021) studied the aqueous and  
393 methanolic extract from three species of pitaya fruits, *Hylocereus costaricensis*, *Hylocereus*

394 *undatus* and *Hylocereus megalanthus*. and obtained an activity of 17 and 13  $\mu\text{mol TE/g}$  sample,  
395 15.3 and 13.2  $\mu\text{mol TE/g}$  of sample and 16.6 and 14  $\mu\text{mol TE/g}$  sample, respectively.

396 As for  $\text{ABTS}\cdot^+$  radical capture, the antioxidant activity of the acetone extract showed  
397 higher activity (829.09  $\mu\text{mol TE/g}$  sample) compared to the other extracts (e.g., aqueous 90 °C  
398 > ethanol > methanol > 45 °C aqueous > 25 °C aqueous). Rufino, Alves, de Brito et al. (2010)  
399 analyzed the combined extract of methanol and acetone of *Euterpe oleracea*, *Copernicia*  
400 *prunifera*, and *Euterpe edulis*, three fruits of the Arecaceae family. They found an activity of  
401 15.1, 10.7 and 78.3  $\mu\text{mol TE / g}$  sample, respectively.

402 Based on the results, it can be noted that the extracts studied showed a relationship  
403 between phenolic compounds and antioxidant activity, once those with the highest  
404 concentration of polyphenols were the ones with the highest antioxidant activity regardless of  
405 the method used.

406 The cytotoxic concentration, i.e., the amount of a product (natural or synthetic) that is  
407 toxic to a given cell, must be determined in order to establish a safety margin for use in  
408 products, taking into account their biological activities (Santos et al. 2013). Figure 4 shows the  
409 results of the cytotoxicity assay by means of the MTT assay to analyze the cell viability of the  
410 L-929 (A) and Caco-2 (B) cell lines in the 90 °C aqueous, ethanolic and acetonic extracts of  
411 the *Aiphanes aculeata* fruit pulp. They were selected for this experiment on the grounds that  
412 they showed a better antioxidant activity. The method has the principle of evaluating cell  
413 viability based on damage induced in mitochondria, which are quantified by reducing MTT (a  
414 yellow-colored salt in water) to formazan (purple-colored crystals, insoluble in water). The  
415 reduction of MTT to formazan is directly proportional to mitochondrial activity and cell  
416 viability (Li & Song, 2007).



418 **Fig. 3.** Effect of *Aiphanes aculeata* extract treatment on L-929 (A) and Caco-2 (B) cell  
 419 viability. The effect of the aqueous extract at 90 °C, (Et) ethanol extract and (Ac) acetic  
 420 extract, at concentrations of 31.25-500 µg/mL on cell viability were evaluated using the MTT  
 421 assay. Values are expressed as mean ± standard deviation (n = 3) and are representative of three  
 422 independent experiments with similar results. \*\*\*p < 0.001, significant difference compared  
 423 to CN (one-way ANOVA, post hoc Tukey). There were no significant differences between  
 424 groups (two-way ANOVA, Bonferroni test), CN (Negative Control).  
 425

426 Cytotoxicity assessment using non-tumor cell lines is an important tool for risk  
 427 prediction and safety assessment of the potential toxicity of bioactive compounds. *In vitro*  
 428 systems are used primarily for screening purposes, for example, to assess cell selectivity and  
 429 to establish more comprehensive toxicological profiles (Eisenbrand et al. 2002). A sample is  
 430 considered cytotoxic when cell death occurs and the percentage of viability of exposed cells is  
 431 < 70 % (Fraga et al. 2021). Therefore, the more toxic the compound, the lower the cell viability.

432 When evaluating the extracts using ethanol and hexane solvents from the *Talisia*  
 433 *esculenta* fruit pulp and peel using the L-929 cell line to evaluate the cytotoxic potential of the  
 434 extracts, Fraga et al. (2021) determined that there is no cytotoxic potential at both tested  
 435 concentrations (12.5 - 200 µg/mL) in relation to the negative control, which are cells without  
 436 any treatment, since there was no cell death, with cell viability above 70 %.

437 Having this in mind, the extracts were evaluated by 2 cell lines, being the fibroblasts L-  
 438 929 non-tumor cells and Caco-2 tumor cells, aiming to evaluate their behavior and selectivity  
 439 against different cell lines. Regarding the L-929 lineage, cell viability was higher than 70 % at

440 all concentrations of both extracts, with no significant difference compared to the control  
441 condition ( $p < 0.05$ ), showing no cytotoxic potential. However, for the Caco-2 cells, the  
442 ethanolic and acetic extracts showed cytotoxic potential at the concentration of 250  $\mu\text{g/mL}$ ,  
443 even decreasing cell viability at the concentration of 500  $\mu\text{g/mL}$  (20.43, 15.13 and 24, 26 %)   
444 for the aqueous 90 °C, ethanolic and acetic extracts, respectively. Pásko et al. (2021) reported  
445 a cell viability of 31.7 and 51.6 % when evaluating the cytotoxicity of aqueous and methanolic  
446 extracts of the *Hylocereus costaricensis* fruit pulp in Caco-2 cells at a concentration of 500  
447  $\mu\text{g/mL}$ , respectively.

448

#### 449 **4. Conclusion**

450 The present study proved to be of great importance as the *Aiphanes aculeata* fruit pulp  
451 presented a rich and varied nutritional and mineral composition. Concerning the total phenolics  
452 content and antioxidant activity, the results indicate a strong correlation, since the aqueous 90  
453 °C and acetic extract showed a greater antioxidant activity in the methods performed. The  
454 active concentrations of these extracts did not show cytotoxicity to fibroblast cells. There were  
455 no significant differences between the extracts and the negative control (untreated) in the entire  
456 concentration range tested. This result is positive evidence for the extracts to be used in food  
457 formulations with functional properties. Regarding the human colon adenocarcinoma cell line  
458 (Caco-2), the two highest extracts concentrations significantly decreased cell viability.  
459 Nevertheless, further studies are needed, or even different forms of extraction, in order to  
460 isolate and evaluate possible compounds with antitumor potential. The results from the present  
461 study demonstrate the importance of a proper exploration and knowledge of native species, as  
462 many have great nutritional and economic potential. However, they remained to be  
463 underexplored.

464

465 **Conflict of interest**

466 The authors declare that they have no conflicts of interest.

467

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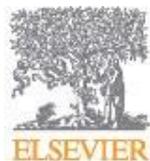
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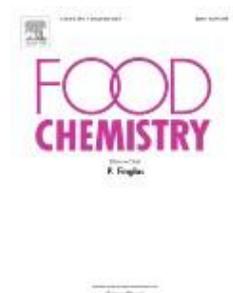
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## DESCRIPTION

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*Food Chemistry* has two open access companion journals [Food Chemistry: X](#) and [Food Chemistry: Molecular Sciences](#).

**The Aims and Scope of *Food Chemistry* are assessed and modified on an annual basis to reflect developments in the field. This means that research topics that have been deemed in scope previously may now fall outside of the scope of the journal as our scientific and technical understanding of the fields evolve and topics become less novel, original or relevant to *Food Chemistry*.**

*Food Chemistry* publishes papers dealing with the advancement of the chemistry and biochemistry of foods or the analytical methods/approach used. All papers should focus on the novelty of the research carried out.

Research advancing the theory and practice of molecular sciences of foods or cure/prevention of human diseases will not be considered for inclusion in *Food Chemistry*.

Topics featured in *Food Chemistry* include:

- Chemistry relating to major and minor **components of food**, their nutritional, physiological, sensory, flavour and microbiological aspects;

- **Bioactive constituents** of foods, including antioxidants, phytochemicals, and botanicals. Data must accompany sufficient discussion to demonstrate their relevance to food and/or food chemistry;
- Chemical and biochemical composition and structure changes in molecules induced by processing, distribution and domestic conditions;
- **Effects of processing** on the composition, quality and safety of foods, other bio-based materials, by-products, and processing wastes;
- Chemistry of **food additives, contaminants**, and other agro-chemicals, together with their metabolism, toxicology and food fate.

Analytical papers related to the microbiological, sensory, nutritional, physiological, authenticity and origin aspects of food. Papers should be primarily concerned with new or novel methods (especially instrumental or rapid) provided adequate validation is described including sufficient data from real samples to demonstrate robustness. Papers dealing with significant improvements to existing methods, or data from application of existing methods to new foods, or commodities produced in unreported geographical areas, will also be considered.

For Analytical Papers, especially those dedicated to the development and validation of methods, authors are encouraged to follow internationally recognized guidelines, such as EURACHEM - for chemical compounds (<https://www.eurachem.org/index.php/publications/guides/mv>) or FDA - for microbiological data (<https://www.fda.gov/downloads/ScienceResearch/FieldScience/UCM298730.pdf>) and proper statistical methods should be applied. Special attention should be given to linearity, selectivity, determination of LOD/LOQ, repeatability and reproducibility of the analysis. Authors should also pay attention to trueness and, when possible (quantitative methods), determine the uncertainty of measurement. Overall, real samples should be analyzed by the state-of-the-art and the newly developed method for validation purposes.

- Methods for the determination of both major and minor components of food especially nutrients and non-nutrient bioactive compounds (with putative health benefits) will be considered.
- Results of method inter-comparison studies and development of food reference materials for use in the assay of food components;
- Methods concerned with the chemical forms in food, nutrient bioavailability and nutritional status;
- General authentication and origin [e.g. Country of Origin Labelling (COOL), Protected Designation of Origin (PDO), Protected Geographical Indication (PGI), Certificate of Specific Character (CSC)] determination of foods (both geographical and production including commodity substitution, and verification of organic, biological and ecological labelling) USING CHEMICAL MARKERS, providing sufficient data from authentic samples should be included to ensure that interpretations are meaningful.

*Food Chemistry* will not consider papers that focus on purely clinical or engineering aspects without any contribution to chemistry; pharmaceutical or non-food herbal remedies; traditional or folk medicines; or survey/surveillance data.

Papers on therapeutic application of food compounds/isolates for treatment, cure or prevention of human diseases will not be considered for inclusion in *Food Chemistry*.

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Food technologists, scientists and chemists

## IMPACT FACTOR

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## ABSTRACTING AND INDEXING

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**Owen Jones**, Purdue University, West Lafayette, Indiana, United States of America

**Mun Yhung Jung**, Woosuk University Department of Food Science and Biotechnology, Wanju-gun, South Korea  
**Simon Kelly**, Food and Environmental Protection Laboratory | Joint FAO/IAEA Centre of Nuclear Techniques in Food and Agriculture | Department of Nuclear Sciences and Applications | International Atomic Energy Agency | Vienna International Centre, Vienna, Austria

**Brad Kim**, Purdue University Department of Animal Sciences, West Lafayette, Indiana, United States of America

**Hua Kuang**, Jiangnan University, Wuxi, Jiangsu, China

**Jungmin Lee**, USDA-ARS Horticultural Crops Research Unit, Corvallis, Oregon, United States of America  
**Hetong Lin**, Fujian Agriculture and Forestry University, Fuzhou, China  
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**Ashkan Madadlou**, National Research Institute for Agriculture Food and Environment Bretagne-Normandie Center, Rennes, France  
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**Jaе-Han Shim**, Chonnam National University, Gwangju, South Korea  
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**Rong Tsao**, Agriculture and Agri-Food Canada Guelph Research and Development Centre, Guelph, Ontario, Canada  
**Shujun Wang**, Tianjin University of Science and Technology, Tianjin, China  
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## GUIDE FOR AUTHORS

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*Journal of Scientific Communications*, 163, 51–59.  
<https://doi.org/10.1016/j.sc.2010.00372>.

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